

Newsletter No. 265 – June 2011

Annual General Necting

Our Next meeting, the June meeting, will be the AGM. By chance, a good number of the present committee will be absent in July, so it was decided to bring the AGM forward to June 21st. All committee positions will become vacant and we will seek nominations for President, Vice-President, Treasurer, at least three General Members, and newsletter Editor. Please give some consideration to making yourself available for a committee position.

Photo Competition

Our now traditional photo competition always accompanies the AGM so it, too, has been moved to the June meeting. Categories this year are Australian Wildlife, Australian Plants, Australian Landscapes, and a new category - Funny Photos. The photo competition is always great fun, with prizes for each category, so bring your fabulous photos to share with the other members on June 21st.

Subs for 2010/2011

The committee has decided that subs must increase for the coming year. The current fee structure is almost pre-historic and does not realistically reflect the costs incurred by the club on your behalf. A number of factors were taken into account in setting the fee levels.

Given the current insurance climate we can no longer accommodate non-members on club outings, excursions and functions. We encourage all of you to take up a family membership to ensure that your wife/husband/children/significant others can attend, and we have structured the fees to make this affordable.

The newsletter is quite costly to produce and mail, so we encourage all members to receive it via email, at no cost to you or the club. We do appreciate that not all of you want to get your '*Correa Mail*' electronically, but there is a cost which must be met by those who choose the mail option. Fees for 2010/2011 are:- Single Member \$15, Family Membership \$20, both of which include the Correa Mail delivered electronically. If you wish to have the newsletter mailed, there will be a fee of \$15 above the membership fee to cover the costs involved.

Pre-meeting Dinner

Some members get together for dinner before the monthly meetings. Dinner before the AGM will be at Mexican Graffiti in Yarra Street, Geelong at 6.00 pm. Discussion will take place at the dinner to decide on future venues.

May Meeting

Orchids

Our speaker was Neil Anderton, who gave us a detailed and interesting talk on his work of cultivating rare terrestrial orchids for reintroduction to the bush. Orchid habitat has been decreasing steadily since European settlement, and some orchid need a helping hand to continue to survive.

All terrestrial (ground growing) orchids have a symbiotic relationship with mycorrhizal fungi which assist the orchids to obtain certain nutrients from the soil. Without these fungi, orchids will not grow. So in order to cultivate orchids from seed, one must first cultivate the fungus.

Luckily the fungus resides in the collar and root system, so samples are readily obtainable. But, obtaining them is a delicate procedure so as not to kill the plant. A hole is made in the soil about 100mm from the plant and then carefully excavated closer, so that the root system isn't damaged. Once the base of the orchid is encountered, the roots are washed with pressurised water and the sample is taken. This can be either a tiny slice from the collar, or a small lateral root, depending on the species of orchid.

The root sample is taken to the lab, thoroughly washed, sterilised in a bleach solution, and washed again. Using a microscope, the fungal balls, called peletons, are removed from the sample, placed in a drop of sterile water and transferred to a petri dish with a mixture of nutrients in agar, to optimise fungal growth. In one or two weeks, the fungus is transferred to another dish and allowed to grow for six weeks at room temperature, then placed in sterile water for long term storage.



A Leek orchid ready for taking a sample

Orchid seed is collected by the quaint method of wrapping the green seed head in an emptied-out teabag, and allowing the seed to ripen. The collected seed is dried and stored at -20°C. When ready, the seed is sterilised, washed, and placed onto a filter paper. This is transferred to a petri dish with a nutrient media suitable for the mycorrhizal fungal growth. Small portions of the fungi are introduced, and the dish is wrapped in foil for 4–12 weeks to simulate below ground conditions.

After about 6 weeks, the seedlings are transferred to a sterile vermiculite and nutrient medium to optimise seedling growth. After a further 2 - 6 months, the seedlings are large enough to be deflasked into pots in non-sterile (natural) conditions.



Seedlings ready to be transferred to vermiculite

They are kept in a mist tent, where automatic misting sprinklers maintain a high (95%) relative humidity. After an appropriate growing time, the seedlings are moved to the growth house where they are tended until they can be rehabilitated into ' the wild'.

Terrestrial orchids can be grown successfully in pots at home. They can be purchased commercially as

seedlings or tubers, but this can be an expensive proposition, By joining the Australian Native Orchid Society (ANOS), orchid tubers can be purchased from the societies tuber bank, or pots are available for sale at the monthly meetings. Terrestrial orchids require a loose, open potting mix which drains well. More orchids are lost from over-watering, than from underwatering. They require minimal or no fertilising, and phosphate is particularly harmful to them. Potting is usually undertaken in December or January.

Tubers can be planted by placing them on top of the potting mix, then layering another 30mm on top, or making a hole and carefully dropping the tuber in. Water them with a fine spray and mulch with chopped casuarina needles. Place the pots in a well lit area and water every 8 - 14 days, until the plant begins to wither, at which time watering ceases. Once the plant has withered and tubers are dormant, they can be repotted, if required.

Our thanks goes to Neil for a wonderfully informative talk, for the use of his photos in this article, and for his on-going work conserving orchids for future generations.



Terrestrials on display at the ANOS (vic) Annual Show

Plant Table

The plant table this month featured some very interesting plants. *Hakea clavata* with its club-like, almost succulent leaves and rich pink flowers solicited a lot of comment, as did the autumn flowering form of our national floral emblem, *Acacia pycnantha*. There were several Eremophilas, which seem to have enjoyed the extra rain of late, and two delightful Diplolaenas - *D. angustifolia* and *D. grandiflora*.

But the table was dominated by the Banksias. Banksia bauera, the possum banksia, features huge flowers. Our specimens were 200mm high and 120mm in diameter. The form of the banksia flowers is more or less constant, but the colours are quite varied. *Banksia spinulosa* has a lovely gold and red flower, while *B. prionotes* is a stunning orange and white. The lovely compact yellow and brown B. media contrasted with the 200mm long, golden yellow flowers of Banksia 'Giant Candles'. We also had specimens of *B. menziesii, B. attenuata,* and *Banksia quercifolia,* which Marg Grabert chose as the plant of the month for July.

Plant of the Month

Banksia quercifolia

by Brendon Stahl

Banksia quercifolia, the Oak-Leaved Banksia, is found in Western Australia near the south coast, from Windy Harbour to Cheyne Beach in the Albany area. It grows in sand, often peaty, in depressions and on swamp margins, in shrub land /sedge formations, sometimes in low woodland. Annual rainfall of the area is 800 – 900 mm. It flowers in the autumn and early winter, from April to July.

In cultivation, it is a fast growing bushy shrub to about 3 metres. If grown from seed, it will, flower in 3 -5 years. The oak-like leaves are bright green on orange/red stems. The cylindrical flower spikes are about 100 mm long by 60 mm wide, greyish in bud and opening to a rusty orange/brown. The plant tolerates medium pruning, but not below green foliage.

It likes a sunny positions or half shade, in well drained or winter moist soil. *B.quercifolia* is one of the few banksias that grow naturally in low lying areas and so may tolerate moisture more than other species.



Banksia quercifolia - photo M Denton

At Deans Marsh, I have one bush more than 17 years old, planted by the previous owner, and some younger plants that I have propagated from seed. I believe it is an attractive shrub.

Australian Garden Wins at Chelsea

A garden featuring Australian native plants has won the gold at the 2011 Chelsea Flower Show. The work of a team from the Melbourne RBG, the garden takes the visitor on a journey, following water's progress, from the red centre to the populated east coast. It features a host of Australian plants, form mature trees, flowering shrubs and rare or unusual plants. Many have been propagated from seed in the UK, to have them ready for the flower show in the northern spring. But the final result was not without its problems.

Mature trees came from nurseries in Italy and Spain and were too tall to be taken into the greenhouses for protection against an unseasonably cold Somerset winter. So they had to be wrapped, laid on their side, and turned regularly. Some of the seedlings were sheltering indoors from the weather, and were browsed by the nurseryman's cat.

Designer Jim Fogarty based the garden on the multiaward winning Australian garden RBG Cranbourne, which is currently undergoing major reconstruction.



Photo – Sydney Morning Herald.

The Chelsea Gold winning garden features a fire pit, "salt" sculptures made from white granite, tonnes of red 'outback' earth and a beautiful billabong in the shape of a boomerang. The garden tells the story of the complex journey taken by Australian river waters, through the outback and its diverse flowering plants, past the saltpans, waterholes and dry riverbeds, into and through the artesian basin and on to the coast. The water reappears as a cascade down a rusted steel gorge and disappears once again into underground aquifers to begin its travels once more.

Up-coming Events

RBG Cranbourne Plant Sale21st AugustSunday 21st August. Open to the general public from9.30am - 4.00pm with prices ranging from \$3.00 -\$6.00 per plant. The nursery is in the Depot area ofthe gardens right next to the main carpark.

Wider Geelong Flora Lecture

Hosted by the Geelong Field Naturalists, the speaker will be Claire Moxham, whose topic is - 'Understanding Coastal Moonah Woodland'

14th June

Coastal Moonah Woodland is listed as a threatened plant community under the *Flora and Fauna Guarantee Act 1988*. Much of the community has been cleared or fragmented, leaving remnants that have been degraded by weed invasion and recreational pressures. Coastal Moonah Woodland is in a demonstrable state of decline, which could result in its extinction if it remains unchecked. Currently it covers less than 10% of its original distribution.

The publication of *A field guide to Coastal Moonah Woodland in Victoria* marks the completion of a six year collaborative research program undertaken by scientists at the Arthur Rylah Institute for Environmental Research (ARI/DSE). Many individuals, local community groups, land managers and government agencies have co-produced this new knowledge about ecological dynamics of the threatened plant community Coastal Moonah Woodland, to better inform management decisions.

Since 2009, Claire has been leading research in the Vegetation Monitoring program at Arthur Rylah Institute for Environmental Research through landscape scale, multi-catchment, partnership projects that foster strategic ecological solutions leading to enhanced biodiversity and production outcomes. Claire's research is focused on various aspects of vegetation ecology and ecosystem function to enable improved management practices to enhance biodiversity conservation, with particular focus grasslands, grassy woodlands and mallee ecosystems.

The lecture is at the Geelong Botanic Gardens Meeting Room. Arrive 7.00 p.m. for a chat & cuppa. Lecture starts at 7.30 p.m.



In Search of *Grevillea kennedyana* Article and photos by Roger Wileman.

In October 2010 I talked to Matt and Pam Baars about their recent trip to Cameron's Corner and the Sturt National Park. They told me how the recent rains had transformed the area into a green paradise with wildflowers everywhere and huge numbers of different species of birds. I did not need any more convincing to go, so after a few phone calls, fuelling and packing the 4wd, five of us, in two vehicles, were off.



One of our main objectives was to find *Grevillea kennedyana*, which we hoped would be in flower. *Grevillea kennedyana* was named in honour of Mrs M.B. Kennedy who collected specimens for Ferdinand Von Meuller in the North West of N.S.W. This grevillea is very rare in cultivation and, as far as I know, is the only one growing wild in three states as it occurs at the junction of the NSW, SA and QLD borders.

Grevillea kennedyana is a well formed plant 1.5 m in height with silver grey , prickly foliage and bright red spider flowers – a very attractive plant. We didn't have any luck finding it at Cameron's Corner but we did find a lot of cowboys in the pub with hats bigger than Frank's. It made me wonder if their necks were strong enough to support the hats.

Back to Tibooburra and, armed with a verbal mud map from the lady at the rangers office, we headed out 80km to the Sturt National Park, where she had seen a plant with red flower half way up a "mesa". We travelled a long way out until we entered the "moonscape" area and further on into the "nuclear" area at the start of the Breakaways, where the land rises up to a high plateau.



Sure enough, right where the lady at the ranger's office said it would be, we found 50-60 plants. Really, they weren't very hard to find as they were the only plant above knee height. *Grevillea kennedyana* is a

very attractive plants and is something that should be in cultivation.

We travelled in the luxury of 4WD vehicles but to think this plant was discovered by W. Baeuerlen in 1887, makes one wonder what he doing was out there.



Further Notes on *G. kennedyana* by Tony Cavanagh

We saw a plant of this species (which I had never previously heard of) at the Myall Park Botanic Garden near Glenmorgan during a recent trip to Queensland. My first reaction was "what an ugly plant", as the picture perhaps shows.



Grevillea kennedyana

Photo-Liz Cavanagh

It is claimed to be "vulnerable" although Olde and Marriott state in *The* Grevillea *Book* that a population of some 7000 had been located, presumably in the 1990s. The plant at Myall Park is grafted onto *G. robusta*, just one of several rootstock tried on an effort to bring this plant into cultivation. As Roger's pictures and story show, it grows in pretty inhospitable country and is very rarely cultivated. The second picture shows a few flowers (nothing like the huge clusters in Roger's pictures) but most interestingly, clearly shows the short, dagger-like silvery "galvanised" leaves which contrast so well with the deep red flowers. Why do such interesting plants have to be so prickly?



Grevillea kennedyana Photo-Liz Cavanagh

Bendigo Bus-trip

The day started out cold and gloomy, and remained so until late afternoon, when it became cold, gloomy and dark. We left the Fyansford car-park promptly at 8.30am, with Harry sprinting to catch the bus as it pulled out. We picked up Arthur, Linda, David and Carmel from Lovely Banks, and headed into the wilds of Central Victoria. Our destination? – The Goldfields Revegetation Nursery at Mandurang. Our Mission? to fill a 5' x 7' trailer with native plants, and make it back alive. A quick stop on Switch Road, to see the lovely little Brisbane Ranges *Grevillea rosmarinifolia*, a few tiny mosquito orchids, and some lovely fat *Correa reflexa*, and it was back in the bus, urged on by Denise who hadn't had any caffeine for two whole hours.



The bus in the bush - Photo Sheila Deakin

We made it to Daylesford without getting lost very often, then foolishly took Carmel's advice and drove the 25 seat bus and trailer into an off-street car-park that was easily big enough for a mini minor. We wandered the quaint little town in search of hot coffee, then stopped the traffic while Ade backed the trailer (with consummate ease) back onto the main street. Then it was on to Castlemaine to visit Munro Court, once a council depot, and now an oasis of sustainable housing amid a delightful bush garden setting.



Wandering Munro Crt – photo Penny Foster

The brief for the designers was for modern houses that are sensitive to the environment. All eight houses are designed by one architect and the garden too, is designed by just one person, allowing for continuity of style throughout. The landscape designer brought in over 400 cubic metres of local soil to create mounds on the flat block that would offer privacy to individual houses and create visual interest in the garden. There are no fences between the blocks, and the houses seem to disappear into the landscape, rather than dominate it as in so many suburban developments. It was a most extraordinary experience, and well worth a wander if you are in the area. Back on board, Roger high-jacked the bus and forced the driver to take them to the Butterfly World nursery, where the hostages half-filled the trailer with natives and a stunning array of orchids.



Lunch – photo Penny Foster

We motored into Bendigo without further complications and stopped for a healthy lunch of

barbeque, hot donuts and red wine at Lake Weeroona Park.

By three o'clock we were underway again, and thanks to Roger's GPS-like navigational skills, we made it to the nursery by the scenic route via Sydney. We were greeted by such a staggering array of plants, that it was hard to know where to start, and even harder to know when to stop. By 4.30 the trailer was groaning under its load of plants – mission accomplished!

So it was off to Ballarat, with multiple navigators, giving myriad directions. Finally Liz took them all in hand and we made it to the Inn of Khong Malaysian/Thai/Chinese restaurant, where we stopped for dinner.



Harry gives Diane some advice – photo Penny Foster

The noise level in the bus increased after dinner and the crowd had become somewhat rowdy. With the CD player pumping out the Everly Brothers, and Arthur planking on the back seat, we chorused our way down the highway and home. A wonderful day was had by all. Thanks to Diane and Roger for organising the various stops along the way. I think I speak for us all when I say "Bring on the next one"

What's in the Bush? Photos by Ade Foster

Spiders

This autumn has seen the return of the spiders, and the bush is alive with them. If you think I sound excited by that ... you're right. For the past few years, the dry winters and springs have reduced the number of flying insects, and so many of the baby spiders, denied their natural prey, have not made it to adulthood. But, not this year! At Anglesea, in the Brisbane Ranges, and elsewhere, the orb-weavers are especially prolific.

Frank Scheelings and I had a couple of days at the Grampians over Easter, ostensibly orchid hunting. However, it was so difficult to walk in the usual 'eyesdown' fashion, because we were constantly bumping into the silken snares of the orb-weavers. The orchids were few and the spiders many, so we turned our attention, and cameras, to them.



Nephila ornata with tiny male in attendance

The most noticeable was the Golden Orb Spider, Nephila ornata. The females are large spiders with a leg span of about 10cm. The bulbous abdomen is brownish, but covered in a smooth pelt of silvery hairs. The legs are brown with reddish joints, silvery hairs and black hair tufts. Their common name derives from the glorious golden colour of the silk in their elaborate webs. Like all the orb-weavers, they spin a typical spoked-wheel web, in which they hang, day and night, in a head down attitude.

But the webs are protected by a series of guardlines which are strung at angles a short distance from the webs. These are probably early warning devices for the spider – touch one and they scamper up an anchor line to put themselves out of harm's way. This silk is remarkably strong and, with the current profusion of spiders, they interlink, making a stroll through the bush almost impossible. These large spiders are all females, and they are attended by several tiny males, who inhabit the outer edges of the web waiting for an opportunity to mate.



Garden Orb Spider – Eriophila biapicata

There were a host of others too. The Garden Orb Spider, *Eriophila biapicata*, was very common and approached the Nephilas in size. They are a greybrown, quite hairy spider with beautiful red, spiky legs. They usually rest in the nearby foliage during the day, with their legs tucked in, and are beautifully camouflaged. Why this population chooses to sat in the webs is a mystery. We also encountered the Sixspined Spider, *Austracantha minax*, the Leaf-curling Spider, *Phonognatha graffei*, and host of other smaller orb spiders.

One of particular interest was the little spider, Cyclosa. They spin a small orb web, with the remains of previous meals arranged in a vertical, silk wrapped larder in the centre. This is common practice with many of the orb weavers. But *Cyclosa* at reast so closely resembles one of these pieces of detritus, that she can sit in her larder, in full view, while remaining almost completely invisible. The abdomens of *Cyclosa* are extended into a variety of shapes, helping to distract the eye of a predator which may be seeking a 'typical' spider.



Cyclosa sp – Photo Frank Scheelings

Next time you are in the bush or the garden in autumn, keep an eye out for the spiders. Once you over-come your revulsion, which is a completely irrational, learned response, it's really quite rewarding. Trust me!

The best time for *Eremophila* cuttings? Article and photos by Tony and Liz Cavanagh.

Over the years, we have taken cuttings at various times of the year – in spring, after plants have finished flowering, when they have "good cutting material", and for the last two years, in summer. Last year, it was in mid January, this year between the 7th and 10th of February. This year's *Eremophila* cuttings were by far the most successful we have ever taken, with roots

appearing in 2-3 weeks and 100% strike for so many species, a success rate we have not had before. Other (non *Eremophila*) species don't appear to be as satisfactory at this time of year, we guess it is probably generally too hot for them, but Eremophilas (the name means "desert loving") thrive in the warmth. In this article, we will discuss our propagation techniques and compare this year's results with last year's. We will also offer our theory as to why this year appears to have been so good.



Eremophila maculata – Photo Ade Foster

Cutting materials and propagating mixes

We always try to take fresh material in the early morning and process it that day. Always choose healthy, strong growing plants if possible and take your cuttings one to two weeks after good rain if you can. We stand the cuttings in water in an ice cream container, keeping each species together with a rubber band. For Eremophilas, you need "firm medium growth" stems, ie stems which are not too woody and will spring back when bent. A good length is 10 -15 cm (4 - 6 in). There are several cutting mixes available, the commonest today probably being 1 part peat moss to 4-5 parts of perlite. The mix has to be well drained but retain enough moisture to support the cuttings. Being from the "old school", we prefer coarse, washed sand and have found that compressed coir (coconut fibre) when expanded and mixed in the same ratio as for peat moss/perlite, is very satisfactory. It is important that you thoroughly wash any sand you intend to use to remove fines and then sterilise it by boiling it for 10-15 minutes. The beauty of the sand mix is that you can reuse it. What's that, reuse propagation mix? Yes you can, simply wash the old mix in a small container and float off all the fines. Boil it for 15 minutes, drain off the water and let the sand cool, and then mix a new batch. We are still using mixes which contain sand we got off the beach at Point Roadknight 20 years ago.

Strip the leaves off at least the lower third of the stem and even though the cuttings may have been taken

only an hour or so previously, always make a new cut at an angle just below a leaf node. Liz uses a scalpel, I prefer sharp secateurs. Dip the cut end in rooting hormone (we prefer Clonex Purple but there are many others), drain for a short time and then insert in the mix. We have found that metal kitchen skewers are ideal for making a hole into which to insert the cuttings. We use "community pots", small black pots 7 cm long by 5 cm diameter, and generally put between 6 and 10 cuttings in a pot. However, hairy and woollyleaved species such as E. subfloccosa need wider spacing, perhaps 4-5 to a pot, as they are very susceptible to fungus and rot and need to be kept somewhat drier than other species. Make sure you label and date each pot. We use a small glasshouse on a sand base to hold the pots. This faces north and is covered with removable 70% shade cloth which is left off on dull or cloudy days. It is vitally important to check cuttings daily, more frequently in hot weather, and water only when needed. A pot of cuttings only needs to dry out once and it is finished. The mix needs to be moist at all times but not too wet, and avoid watering the foliage as much as possible.

The most exciting time with growing from cuttings is when roots begin to appear at the base of the pot. As some species produce small, weak roots, do not ever be tempted to check whether cuttings have rooted by pulling on the stems. Patience is a virtue. This was not a problem for us in 2011. Liz found roots on some species before the end of February and I did the first potting up on 3 March – just 25 days – and a major one on the 14th. The picture below shows just how massive the roots were and more amazingly, nearly every pot had a 100% strike.



Massive roots on E. sargentii

Sometimes, one or two cuttings will have little or no roots while the rest in the pot are well developed. These can be reset and will often root some time later. When cuttings produce such dense roots, it is often difficult to separate them and this leads to the question "how is the best way to remove rooted cuttings from a pot?" We always water the cutting pots first and then gently knock the mix from the pot before immersing it in a bucket of water. This method allows you to recover the sand but more importantly, the tangled roots can be separated under water with minimum damage and the individual cuttings can then easily be potted up. Make sure that the roots don't dry out before potting.

This year we were growing a lot of Eremophilas for our daughter who lives in Mildura and were able to compare the results for 2010 and 2011 for many species. Many of the plants are relatively easy to strike but nevertheless, we were fascinated by just how much better this year's results were in relation to 2010. The two years are compared in the table on the next page.

So what does it all mean? Because these were all single sets of cuttings with no duplicates, we can't draw any conclusions about individual results. We can say that 2011 was definitely a much better year than 2010 in that cuttings rooted much earlier (from 3 to 6 weeks) with a very high success rate - in fact the most consistent set of results we have ever had. And they rocketed away in the pots! There is also some indication that species which were slower to root in 2011 ie took 36 to 42 days, were mostly the same ones as were slow in 2010, ie took between 80 and 115 days. Such species include E. calorhabdos, some forms of *glabra* (although the results are somewhat erratic), "Kalbarri Carpet", E. racemosa, E. sargentii, although a couple like E. microtheca, E. weldii and sp. "Kalgoorlie" were among the first to root in 2010. And some species appear to be difficult (slow to root and/or often die in their pots) no matter when they are taken. These include E. decipiens, E. hillii, E. subfloccosa, E. pantonii and oppositifolia, Ε. "Summertime Blue".



Well developed & small roots from stems in same pot



Washing the mix off the roots

We have a theory about why we had such success in 2011 and it's all to do with the weather. Over the summer of 2009-10, we were still in drought with December rainfall of 55 mm and perhaps 20 mm up to the time we took cuttings in mid January. Over summer of 2010-11, while we had only around 42 mm in December, we had received 750 mm for the year and around 175 mm up to 8 February (including 60 mm on 4 Feb.). The plants were literally "jumping out of the ground", growing strongly and with plenty of ideal cutting material. Moreover, we had a relatively cool summer with very few extreme days, and many days of high humidity so the cuttings were easy to manage. Anyway, we were delighted with the results and we hope that it may inspire others to try taking cuttings of at least some species such as Eremophila during summer. It can be very rewarding and with luck, you could be planting them out in autumn.



Potted cuttings in early April

Species	Days to 1 st	% strike 2011	Days to 1 st	% strike 2010*
	potting 2011		potting 2010	
biserrata	25	100	50	50
calorhabdos	36	75	95	55
complanata	36	65	NA	
decipiens	Nil		115	35
decipiens subsp. linearifolia	Nil		NA	
glabra (shrub, orange flrs)	36	100	50	25
glabra (sprawling, red flrs)	40	75	115	40
glabra (prostrate, green flrs)	36	100	NA	
glabra (Murchison River, red fl)	Nil		50	<20
hillii	Nil		NA	
"Kalbarri carpet"	36	100	80	40
sp. "Kalgoorlie"? drummondii	36	90	50	75
laanii	42	50	Nil	
maculata (shrub, large, red)	36	100	NA	
<i>maculata</i> (shrub, yellow)	40	100	Nil	
maculata subsp. brevifolia	NA		50	70
microtheca	36	100	50	70
nivea X drummondii	25	90	50	100
oppositifolia (cream)	Nil		NA	
oppositifolia (pink)	Nil		Nil	
polyclada	40	75	NA	
racemosa	36	100	80	35
"Roseworthy"	42	100	NA	
sargentii	36	100	95	35
subfloccosa	Nil		Nil	
subteretifolia	25	100	NA	
"Summertime Blue"	Nil		NA	
superb	25	100	50	50
weldii	40	85	50	100

* = approximate only

NA = not grown this year

Nil = No result (for 2011, as at 30 March)

Newsletter contributions

Once again, I extend a big thank you to Tony for his continued support of the newsletter, and to Roger who has also contributed an article this month. I hope in the future to receive articles from each and every one of you so that we can all share in your passion for native plants. Surely you are tired of hearing from me by now?? ⁽ⁱ⁾

My Garden

We mentioned in April that we would like members to do a brief talk about their garden, or a favourite plant at a meeting in spring. Frank and I are happy to take photos on your behalf and assist with a computer display on the night. We'd like you all to take part in this night, and we'd love to have it become a regular part of our year. Please give some thought to what you are going to tell us about on the 'My Garden' night.

Membership Forms

Accompanying this newsletter you will find a membership form for the 2011/2012 year. Please fill in ALL the details on this form and return it with your subs. Unless you tell us otherwise, the email address and phone number will only be used when we need to contact you urgently, and both will be treated as private. Thanks for your co-operation.



Australian Plants Society Geelong

Membership Form 2011/2012

Name					
Address					
				Post Code	
Phone					
Email					
Newsletter	•••	By Mail(\$15)	By e	Mail (no cost)	(Please circle)

Membership fees 2011/2012

Single Membership	•••	\$ 15.00
Family Membership	•••	\$ 20.00
Newsletter by mail		\$ 15.00
Total	•••	\$

My payment accompanies this form

Signature

Please mail to APS Geelong, P.O. Box 2012, Geelong 3220 Or hand to the Treasurer